

Effect of aqueous fruit extract of *Xylopia aethiopica* fruit on some fetal growth parameters and histology of uterus in Wistar rats

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ABSTRACT

Introduction: *Xylopia aethiopica* (*X. aethiopica*) fruit is widely used in Africa as a food additive. It is also employed to prevent nausea during pregnancy, aid uterine contractions during childbirth, treat menstrual flow anomalies, uterine fibroids, and malaria. With the increased consumption rate among pregnant women, especially in Nigeria, the present study aimed to evaluate the effects of the aqueous fruit extract of *X. aethiopica* on developing fetuses and pregnancy in rats. **Methods:** Twenty pregnant female rats (gestation day zero) were randomly assigned to four groups of five rats each. Group I served as the control and received distilled water, while Groups II–IV were given *Xylopia aethiopica* fruit extract at doses of 100 mg/kg, 200 mg/kg, and 300 mg/kg, respectively, for 18 days (days 1 to 18 of pregnancy). All rats were euthanized under ketamine injection on day 19. The abdomen was opened, and the number of live and dead fetuses and the crown-rump length were measured. The uterine horns were fixed in 10% formalin for light microscopy. **Results:** Administration of the aqueous fruit extract of *X. aethiopica* at 100 mg/kg, 200 mg/kg, and 300 mg/kg for 18 days significantly reduced ($P < 0.05$) the number of live fetuses, fetal viability, and crown-rump length, while increasing the number of dead fetuses compared to the control. Histological examination of the maternal uterine horn revealed distortion of the uterine epithelium in rats treated with 200 mg/kg *X. aethiopica*, and degenerating uterine epithelium and endometrial connective tissue in rats treated with 300 mg/kg *X. aethiopica*. **Conclusions:** Continuous consumption of *X. aethiopica* fruit during pregnancy could be toxic to the developing fetus and affect maternal health. Hence, caution should be exercised when consuming *X. aethiopica* fruit during pregnancy.

Key words: Fetus, *Xylopia aethiopica*, uterine horn toxic, epithelium

INTRODUCTION

Xylopia aethiopica (Dunal) A. Rich., also known as African or Ethiopian pepper, is a medicinal plant distributed in the lowland rainforests of the Guinea Savannah zones of Africa. It is cultivated in Angola, Ethiopia, Nigeria, Senegal, and Sudan¹. This plant is commonly used in the preparation of African dishes, particularly in Nigeria, and in traditional medicine for managing various ailments such as skin infections, candidiasis, dyspepsia, cough, biliousness, febrile pains, bronchitis, rheumatism, dysentery, and boils^{2,3}. *Xylopia aethiopica* has various pharmacological properties, including analgesic, anti-inflammatory, anti-allergic, anti-allodynic, antispasmodic, anti-hyperalgesic, antidepressant, antioxidant, and hypoglycemic effects^{2,4-7}.

The plant has demonstrated significant progress in reproductive medicine, including hastening fetal delivery, supporting prenatal development, and serving as a pre- and post-coital contraceptive⁸. Herbal medicine is often preferred due to its availability

and a long history of proven effectiveness⁹. The female reproductive system is sensitive to environmental and chemical factors such as lifestyle, radiation, drugs, and toxicants¹⁰. Exposure to these factors may lead to congenital abnormalities in fetuses and affect adult physiology¹¹, potentially impacting reproductive capability¹². Toxicity during pregnancy can exert various effects on fetal development and maternal health¹³. The reproductive and child health program aims to reduce maternal deaths and improve child health outcomes¹⁴. The use of plants for medicinal and traditional purposes to address health issues has been common in African and other societies for centuries¹⁵. Many herbal remedies are traditionally used as contraceptives (to prevent ovulation or fertilization), abortifacients (to prevent implantation), and as emmenagogues (to regulate uterine flow) or oxytocics (to stimulate uterine contractions, particularly to promote labor)¹⁶.

There is growing concern about health hazards associated with the consumption of food additives^{17,18}, which may have teratogenic effects during preg-

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nancy¹⁹. Therefore, with the increasing consumption rate of *Xylopia aethiopica* fruit, especially among pregnant women, studies are needed to determine the health hazards associated with its continuous exposure during pregnancy and its effects on the developing fetus.

MATERIALS AND METHODS

Chemicals

Ketamine hypochlorite injection was obtained from the University of Maiduguri Clinic, while hematoxylin and eosin stains were purchased from BDH Chemical Ltd (Poole, England).

Plant Materials and Extraction

Dried fruits of *X. aethiopica* were bought from Monday Market, Maiduguri, Nigeria. Aqueous extraction was carried out as described by Adienbo et al.⁵. The fruits were pulverized, and 330 g of the powder was soaked in three litres of distilled water. The mixture was refluxed for two hours in a continuous extraction (Soxhlet) apparatus, and the solution was filtered with a thimble to remove debris. The filtrate was concentrated to a powder using a rotary evaporator.

Animals

Forty Wistar rats (20 male and 20 female), weighing between 150–200 g, were used for this study. They were purchased from the National Veterinary Research Institute Vom, Nigeria. The rats were housed in plastic cages at the animal house of the Department of Biochemistry, University of Maiduguri, Nigeria, to acclimatize to the laboratory conditions for two weeks. They were fed with grower mash (Vital Feed, Nigeria) and water ad libitum. The rats were weighed weekly throughout the study.

Experimental Design

The fetal study was carried out according to the method described by Aouni et al.²⁰, with minor modifications. Briefly, adult nulliparous female rats detected in the proestrus stage of the oestrous cycle were cohabited with male albino rats that had not been subjected to any experimental procedures, in a ratio of 1:1 per cage overnight. The presence of spermatozoa in a vaginal smear (identified with the aid of a light microscope) indicated successful mating and, as such, pregnancy was established. That day was considered gestation day zero. Mated females were randomly assigned into four experimental groups of five rats each. Group I served as

the control and received distilled water daily, while Groups II–IV were treated with the aqueous fruit extract of *X. aethiopica* at doses of 100 mg/kg, 200 mg/kg, and 300 mg/kg, respectively. The administration was performed daily, starting from day 1 to 18 of pregnancy. Twenty-four hours after the last administration (day 19 of pregnancy), the rats were euthanized under ketamine injection. The abdomen was opened, and the number of live and dead fetuses, ovarian weight, as well as crown–rump length were measured. The percentage of live fetuses was estimated as the number of live fetuses divided by the total number of fetuses, multiplied by 100. The uterine horns were fixed in 10% formalin for light microscopy.

Histological study

The organs were dehydrated in graded alcohol, embedded in paraffin, and sectioned at 5µm with a rotary microtome (Leica RM2125 Rotary Microtome). The sections were stained with Hematoxylin and Eosin (H&E) and micrographs were taken using a microscope camera (AmScope, UK) at different magnifications.

Statistical analysis

Oestrous cycle and organ index data were analyzed with GraphPad Prism 7 (GraphPad, USA). One-way ANOVA and Dunnett posthoc test were conducted and the results were presented as Mean ± standard error of the mean (SEM). $P < 0.05$ was considered statistically significant.

RESULTS

Maternal Body Weight

According to **Table 1**, there was a non-significant ($P < 0.05$) increase in maternal body weights of rats that were orally administered the aqueous fruit extract of *X. aethiopica* at all doses compared to the control group. Pregnancy weight gains were not altered at any dose level of the extract.

Fetal Growth Parameters

The result from the fetal study (**Table 2**) in the present study revealed that all groups of rats that were orally administered the aqueous fruit extract of *X. aethiopica* at 100 mg/kg (1.54 ± 0.05), 200 mg/kg (1.15 ± 0.03) and 300 mg/kg (0.76 ± 0.00) demonstrated a significant ($P < 0.05$) reduction in crown-rump length compared to the control (2.90 ± 0.05). There was no significant decrease in the mean number of live fetuses of

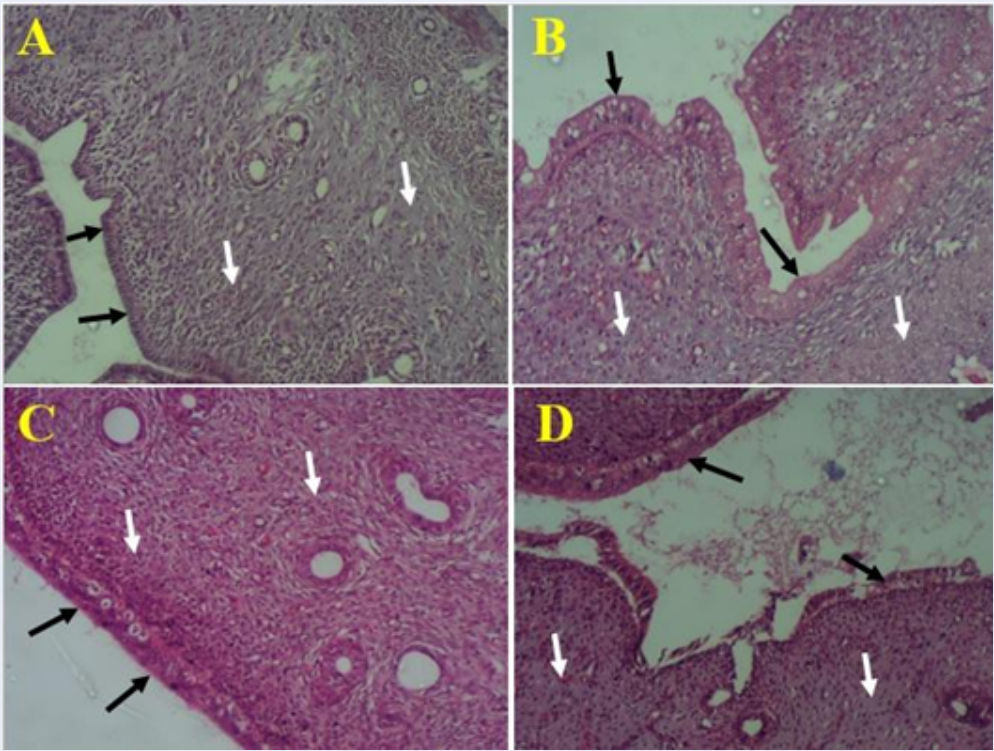


Figure 1: Photomicrograph of uterine horn showing endometrial epithelial lining (black arrows) and connective tissues with glands (white arrows). A= control rats, B= rats treated with *Xylopi aethiopica* at 100 mg/kg, C= rats treated with *Xylopi aethiopica* at 200 mg/kg, while D= rats treated with *Xylopi aethiopica* at 300 mg/kg. H&E stain, x100 magnification.
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Table 1: Effect of Aqueous Fruit Extract of *X. aethiopica* on Maternal Body Weight in Albino Rats

Groups	Dose (mg/kg)	Initial Body Weight (g)	Final Body Weight (g)	Body Weight Differences (g)	Percentage Differences %
Con- trol	0	154.48±9.44	217.76±14.73	63.28	40.96
XA	100	161.98±4.79	179.88±7.88	17.90	11.05
XA	200	150.30±4.11	189.44±17.63	39.14	26.04
XA	300	150.92±4.76	177.38±11.73	26.46	17.53

All values are expressed as mean±SEM (n = 5). * indicates a significant difference with the control at P < 0.05. SEM = Standard error of the mean, XA= *Xylopi aethiopica*.

rats that received *X. aethiopica* extract at all doses when compared to the control. On the other hand, a non-significant increase was observed in the mean number of dead fetuses in the treated groups relative to the control. A dose-dependent decrease was observed in rats treated with fetal viability, there was a reduction in a dose-dependent manner in tested groups, with *X. aethiopica* (25 % in 300 mg/kg) compared to the control (100%).

Histological Observations

The histological examination of the control rats' uterine horn showed normal simple columnar epithelium and extensive lamina propria that bears endometrial glands and myometrium (Figure 1A). Such histoarchitecture of the uterus did not show any visible change after administration of aqueous fruit extract of *X. aethiopica* at 100 mg/kg of the extract (Figure 1B). However, the uterine horn of rats that received 200 mg/kg revealed distortion of the uterine epithelium (Figure 1C) while those of rats

Table 2: Effect of Aqueous Fruit Extract of *X. aethiopica* on Fetal Growth Parameters

Groups	Dose (mg/kg)	Crown Rump Length (cm)	Live Fetuses	Dead Fetuses	Fetal Viability (%)
Control	0	2.90±0.05	6.80±0.37	0.00±0.00	100
XA	100	1.54±0.05*	4.20±1.53	0.40±0.40*	75
XA	200	1.15±0.03*	2.80±1.70*	0.60±0.40*	56*
XA	300	0.76±0.00*	1.00±1.94*	1.00±0.63*	25*

All values are expressed as mean±SEM (n = 5). * indicates a significant difference with the control at P < 0.05. SEM = Standard error of the mean, XA= *Xylopia aethiopica*.

treated with 300 mg/kg of *X. aethiopica* revealed de-generating uterine epithelia and connective tissue (Figure 1D).

DISCUSSION

The present study revealed that the administration of aqueous fruit extract of *X. aethiopica* to pregnant rats caused an increase in maternal mean body weight. This increase may suggest that growth was not impaired by the extract in all experimental groups. Additionally, changes in maternal body weight can be used to evaluate the integrity of maternal homeostasis²¹. This agrees with Onyegeme-Okerenta et al.²², who reported that an aqueous extract of *Millittia* absences increased the body weight of pregnant rats. Although growth was not altered in all treatment groups, the extract still produced varying effects on some fetal growth parameters evaluated in this study. Moreover, the present study showed a non-significant decrease in the weight of the maternal ovaries of rats that received the aqueous fruit extract of *X. aethiopica* compared to the control. The weight of the three endocrine tissues of the ovary—the stroma, follicles, and corpus luteum—constitutes the total ovarian weight²³. Hence, the reduction in ovarian weight in this study may be attributed to the absence of gonadotropin or steroid hormones due to disrupted follicular activity²⁴. Furthermore, this study revealed that administration of aqueous fruit extract of *X. aethiopica* to pregnant rats caused significant growth retardation (decreased crown-rump length), decreased number of live fetuses and fetal viability, and an increased number of dead fetuses in the uterine horns. This could explain the histological observations in the maternal uteri of the groups that received 200 mg/kg and 300 mg/kg body weight of the extract, which revealed distention of the uterine epithelium, reduction in endometrial glands, and distortion of the uterine endometrial epithelial lining compared to the corresponding control. This may result from

disruption of pregnancy by interference with mitotic division of the fetus, causing destruction of the endometrial lining of the uterus²⁵. Likewise, the present findings are in line with reports by Onyebuagu et al.²⁶ and Onyebuagu and Agbai²⁷ on the activities of *X. aethiopica*. They also agree with the study by Aouni et al.²⁰, which documented that hydro-ethanolic extract of *Marrubium vulgare* induces severe histological changes in the pregnant rat uterus. This corresponds with the findings by Zade and Dinesh²⁷ and Choudhary et al.²⁸ on the mechanisms of aqueous leaf extract of *Indigofera trifoliata* and hydroalcoholic leaf extract of *Alstonia scholaris*. Additionally, reduction of endometrial glands is related to infertility and represents a defect in implantation, thus the uterus cannot accommodate a developing fetus to term²⁹. In addition, administration of aqueous fruit extract of *X. aethiopica* caused a reduction in crown-rump length, number of live fetuses, and fetal viability, as well as an increased number of dead fetuses in experimental groups. The study has shown that nearly all drugs administered during pregnancy will, to some extent, enter fetal circulation through passive diffusion^{30,31}. Therefore, since *X. aethiopica* fruit extract was administered during the cleavage and blastula stages of rat embryonic development—before implantation—as well as at later stages of embryo and fetal development, it might interfere with the normal course of pregnancy. This could explain the negative adverse effects observed in the fetal growth parameters in the present study. This agrees with the findings of Garba et al.⁸, that *Cissampelos mucronata* root extract negatively influences reproductive outcomes in pregnant rats. It is also in line with a study by Zade and Dinesh²⁷, who stated that *Indigofera trifoliata* leaf extract (aqueous, alcohol, ethyl acetate, and chloroform) harms developing fetuses. This was equally documented by Yakubu et al.³² on the adverse effects of aqueous leaf extract of *Senna alata* on pregnant rat outcomes. Furthermore, the study revealed that steroids, flavonoids, alkaloids, and phenolics found in a va-

riety of plants have antifertility activity, thereby altering normal fetal growth, development, and pregnancy^{33,34}. Hence, in the present study, saponins, alkaloids, steroids, and flavonoid components of the aqueous fruit extract of *X. aethiopica* might have exerted their antifertility effects, leading to reduced crown-rump length, number of live fetuses, fetal viability, and increased number of dead fetuses in the experimental groups. Similarly, Nwafor and Kalio³⁵ documented that aqueous fruit extract of *X. aethiopica* has strong contractile potential on uterine smooth muscles. This strong contractile property may also contribute to the observed decrease in crown-rump length and number of live fetuses, along with an increased number of dead fetuses in the present study compared to the control. This may cause irritation and stretching of the uterine cervix, subsequently stimulating the posterior pituitary gland to increase its secretion of oxytocin³⁶. Therefore, the presence of alkaloids, flavonoids, and saponins may, in part, contribute to the oxytocic effect of the aqueous fruit extract of *X. aethiopica*³⁷. Since the fruits and seeds of *X. aethiopica* are sometimes added to meals of pregnant individuals to help ease childbirth, there could be health implications for both the mother and the developing fetus if the dosage is not minimized.

CONCLUSIONS

Findings from the present study revealed that repeated consumption of *X. aethiopica* fruit during pregnancy could be toxic to both the developing fetus as well as affecting maternal health. Hence, care should be taken while consuming *X. aethiopica* fruit in pregnancy.

ABBREVIATIONS

ANOVA: analysis of variance, **H&E**: hematoxylin and eosin, **SEM**: standard error of the mean, **XA**: *Xylopia aethiopica*.

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None.

AUTHOR'S CONTRIBUTIONS

All authors conceived and designed the research. KOA & NID provided study materials and conducted the research. All authors analyzed and interpreted the data. KOA & NID drafted the initial manuscript. SHG & JVZ revised the manuscript. All authors approved the final manuscript.

FUNDING

Not applicable.

AVAILABILITY OF DATA AND MATERIALS

The datasets used and/or analyzed during the current study are available from the corresponding author upon reasonable request.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

This study was approved by the Postgraduate Board of Studies, University of Maiduguri, and performed according to the animal research: reporting of in vivo experiments (ARRIVE) guidelines.

CONSENT FOR PUBLICATION

Not applicable.

COMPETING INTERESTS

The authors declare that they have no competing interests.

REFERENCES

- Burkill HM. *Xylopia aethiopica* (Dunal) A. Rich. The Useful Plants of West Tropical Africa Royal Botanical Gardens. 1985;1:11–20.
- Fetse JP, Kofie W, Adosraku RK. Ethnopharmacological Importance of *Xylopia aethiopica* (DUNAL) A. RICH (Annonaceae) - A Review. *Br J Pharm Res*. 2016;11(1):1–21. Available from: <https://doi.org/10.9734/BJPR/2016/24746>.
- Enemchukwu BN, Erimujor SO, Ubaaji KI. Phytochemical Screening and Biochemical Effects of Aqueous Seed Extract of *Xylopia aethiopica*, (Uda) on Selected Haematological Indices in Male Wistar Albino Rats. *The Bioscientist*. 2014;2:103–109.
- Nnodim JK, Emeju A, Amaechi A, et al. Influence of *Xylopia aethiopica* Fruits on some Hematological and Biochemical Profile. *Al Ameen J Med Sci*. 2011;4:191–196.
- Adienbo OM, Nwafor A, Ogbomade RS. Contraceptive Efficacy of Hydro-Methanolic Fruit Extract of *Xylopia aethiopica* in Male Albino Rats. *Int J Adv Biol*. 2013;1:718–727.
- Ameyaw EO, Woode E, Boakye-Gyasi E, et al. Anti-allodynic and Anti-hyperalgesic Effects of an Ethanolic Extract of *Xylopia aethiopica* from the Fruits of *Xylopia aethiopica* in Murine Models of Neuropathic Pain. *Pharmacognosy Res*. 2014;6(2):172–179. Available from: <https://doi.org/10.4103/0974-8490.129041>.
- Okpashi VE, Bayim BP, Obi-Abang M. Comparative Effects of some Medicinal Plants: *Anacardium occidentale*, *Eucalyptus globulus*, *Psidium guajava* and *Xylopia aethiopica* extracts in Alloxan-Induced Diabetic Male Wistar Albino Rats. *Biochem Res Int*. 2014;2014:203051. Available from: <https://doi.org/10.1155/2014/203051>.
- Garba SH, Jacks TW, Onyeyili PA, et al. Embryofetal Effect of the Methanolic Root Extract of *Cissampelos mucronata* A Rich in Rats. *Anat J Afr*. 2014;3:286–293.
- Thomford NE, Dzobo K, Chopera D, et al. Pharmacogenomics of Using Herbal Medicinal Plants on African Populations in Health Transition. *Pharmaceuticals (Basel)*. 2015;8(3):637–663. Available from: <https://doi.org/10.3390/ph8030637>.
- Fucic A, Gamuli M, Ferenic Z, et al. Environmental Exposure to Xenoestrogen and Oestrogen Related Cancers: Reproductive System, Breast, Lung, Kidney, Pancreas and Brain. *Environ Health*. 2012;11(Suppl 1):S8. Available from: <https://doi.org/10.1186/1476-069X-11-S1-S8>.

11. Knapp KM, Brogly SB, Muenz DG, et al. Prevalence of Congenital Anomalies in Infants with Utero Exposure to Antiretrovirals. *Pediatr Infect Dis J*. 2012;31(2):164–170. Available from: <https://doi.org/10.1097/INF.0b013e318235c7aa>.
12. Monima LA, Buhari M, Lawal S, et al. Effect of Cleome gynandra Leaf Extract on the Estrous Cycle and Histology of the Ovary and Uterus of Wistar Albino Rats. *Anat J Afr*. 2019;8(1):1385–1394. Available from: <https://doi.org/10.4314/aja.v8i1.182619>.
13. Auharek SA, Vieira MC, Cardoso CA, et al. Reproductive Toxicity of *Campomanesia xanthocarpa* (Berg.) in Female Wistar Rats. *J Ethnopharmacol*. 2013;148(1):341–343. Available from: <https://doi.org/10.1016/j.jep.2013.04.010>.
14. Tatiya AU, Dande PR, Mutha RE, et al. Evaluation of Saponins from *Sesban L. Merr* on Acute and Chronic Inflammation in Experimental Induced Animal. *J Biol Sci*. 2013;1:123–130. Available from: <https://doi.org/10.3923/jbs.2013.123.130>.
15. Mohammed A, Ibrahim MA, Islam MS. African Medicinal Plants with Antidiabetic Potentials: A Review. *Planta Med*. 2014;80(5):354–377. Available from: <https://doi.org/10.1055/s-0033-1360335>.
16. Ritchie HE. The Safety of Herbal Medicine Use during Pregnancy. *Frontier in Fetal Health*. 2001;3:259–266.
17. Abd-Elhakim YM, Hashem MM, El-Metwally AE, Anwar A, Abo-EL-Sooud K, Moustafa GG, et al. Comparative Haemato-Immunotoxic Impacts of Long-Term Exposure to Tartrazine and Chlorophyll in Rats. *Int Immunopharmacol*. 2018;63:145–154. Available from: <https://doi.org/10.1016/j.intimp.2018.08.002>.
18. Abo-El-Sooud K, Hashem MM, Badr YA, Eleiwa MM, Gab-Allaha AQ, Abd-Elhakim YM, et al. Assessment of Hepatorenal Damage and Genotoxicity Induced by Long-Term Exposure to Five Permitted Food Additives in Rats. *Environ Sci Pollut Res Int*. 2018;25(26):26341–26350. Available from: <https://doi.org/10.1007/s11356-018-2665-z>.
19. Collins TF, Black TN, Ruggles DI. Teratogenic Potentials of FD & C Red No. 3 when given by Gavage. *Toxicol Ind Health*. 1993;9(4):605–616. Available from: <https://doi.org/10.1177/074823379300900403>.
20. Aouni R, Attia MB, Jaafoura MH, et al. Effects of Hydro-Ethanol Extract of *Marrubium vulgare* in Female Rats. *Asian Pac J Trop Med*. 2017;10(2):160–164. Available from: <https://doi.org/10.1016/j.apjtm.2017.01.010>.
21. Almeida FC, Lemonica IP. The Toxic Effect of *Coleus barbatus* B. on the Different Periods of Pregnancy in Rats. *J Ethnopharmacol*. 2000;73(1-2):53–60. Available from: [https://doi.org/10.1016/S0378-8741\(00\)00275-0](https://doi.org/10.1016/S0378-8741(00)00275-0).
22. Onyegeme-Okerenta BM, Anacleto FC, Offor K. Abortifacient Potential Effect of Aqueous Extract of *Millittia aboensis* on Reproductive Health of Matured Wistar Rats. *Journal of Pharmacy and Biological Sciences*. 2016;11:13–19.
23. Rahman SA, Ahmed NA, Samat NH, et al. The Potential of Standardized Quassinoid-Rich Extract of *Eurycoma longifolia* in the Regulation of the Oestrous Cycle of Rats. *Asian Pac J Trop Biomed*. 2017;7(1):27–31. Available from: <https://doi.org/10.1016/j.apjtb.2016.07.016>.
24. Orisaka M, Hattori K, Fukuda S, et al. Dysregulation of Ovarian Follicular Development in Female Rat: LH Decreases FSH Sensitivity during Preantral-Early Antral Transition. *Endocrinology*. 2013;154(8):2870–2880. Available from: <https://doi.org/10.1210/en.2012-2173>.
25. Elbetieha A, Oran SA, Alkofahi A, Darmani H, Raies AM. The fetotoxic potential of *Globularia Arabica* and *Globularia alypum* (Globulariaceae) in rats. *J Ethnopharmacol*. 2000;27(1-2):215–219. Available from: [https://doi.org/10.1016/S0378-8741\(00\)00246-4](https://doi.org/10.1016/S0378-8741(00)00246-4).
26. Onyebuagu PC, Pughikumo DT, Aloamaka CP. Antifertility Effects of Dietary *Xylopi aethiopica* ((Dunal) A. Rich) in Female Wistar Rats. *Nigerian Annals of Natural Sciences*. 2013;13:17–21.
27. Zade V, Dinesh D. Abortifacient Efficacy of *Indigofera trifoliata* Leaves Extract on Female Albino Rats. *Asian J Pharm Clin Res*. 2019;6:75–79.
28. Choudhary M, Rani S, Sharma P, et al. Antifertility and abortifacient potential of hydroalcoholic leaves extract of *Alstonia scholaris* in female rats: an ethnomedicine used by Papua women in New Guinea. *Bull Fac Pharm Cairo Univ*. 2017;2017:55.
29. Adkar PP, Ghadge PM, Ambavade SB, et al. Effect of Hydroethanolic Extracts of Leaves of *Trapa bispinosa* Roxb on Histological Assessment in Reproductive System of Albino Mice. *J Pharm Res*. 2014;8:570–575.
30. Syme MR, Paxton JW, Keelan JA. Drug Transfer and Metabolism by the Human Placenta. *Clin Pharmacokinet*. 2004;43(8):487–514. Available from: <https://doi.org/10.2165/00003088-200443080-00001>.
31. Selderslaghs IWT, Rompay AR, Coen WD, et al. Development of a Screening Assay to Identify Teratogenic and Embryotoxic Chemicals Using the Zebrafish Embryo. *Reprod Toxicol*. 2009;28(3):308–320. Available from: <https://doi.org/10.1016/j.reprotox.2009.05.004>.
32. Yakubu MT, Adeshina AO, Oladiji AT, et al. Abortifacient Potential of Aqueous Extract of *Senna alata* Leaves in Rats. *Reprod Contracept*. 2010;21(3):163–177. Available from: [https://doi.org/10.1016/S1001-7844\(10\)60025-9](https://doi.org/10.1016/S1001-7844(10)60025-9).
33. Oderinde O, Noronha C, Oremosu A, Kusemiju T, Okanlawon OA. Abortifacient properties Aqueous Extract of *Carica papaya* (Linn) seeds on female Sprague-Dawley rats. *Niger Postgrad Med J*. 2002;9(2):95–98. Available from: <https://doi.org/10.4103/1117-1936.171036>.
34. Anderson LL, Maghissi KS, Hafes ES. *Biology of Mammalian Fertilization and Implantation*. Illinois: Thomas Springer-field; 2007.
35. Nwafor A, Kalio ID. Effects of *Xylopi aethiopica* on Pregnancy in Albino Rats. *Journal of Medical and Pharmaceutical Sciences*. 2006;2:1–4.
36. Guyton A. *Guyton Medical Physiology*. 11th ed. London: Saunders Company; 2006.
37. ElMazoudy RH, Attia AA. Ginger Causes Subfertility and Abortifacient in Mice by Targeting Both Estrous Cycle and Blastocytes Implantation Without Teratogenesis. *Phytomedicine*. 2018;50:300–308. Available from: <https://doi.org/10.1016/j.phymed.2018.01.021>.